

# Tyrosine Oxidation by NO<sub>2</sub> in Aqueous Solution

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Nitrogen dioxide, formed by  $\gamma$ -radiolysis in deaerated aqueous nitrate/nitrite solutions, is capable of oxidizing Gly-Tyr in favourable competition with the natural decay of NO<sub>2</sub> by dimerization and disproportionation. 2,2'-Biphenolic tyrosine dimers and nitro-tyrosine were identified spectroscopically as stable products. The results suggest that NO<sub>2</sub> reacts with the peptide by electron abstraction, generating Gly-Tr phenoxyl radicals (PheO<sup>•</sup>) which terminate by dimerization (2 PheO<sup>•</sup> → 2,2'-biphenol) and NO<sub>2</sub>-scavenging (PheO<sup>•</sup> + NO<sub>2</sub> → Nitro-Tyr).

## Introduction

The kinetics of the elementary chemical processes of nitrogen dioxide in aqueous solution have been resolved by application of pulse radiolysis techniques [1]



$$(2k_{1f} = 9.0 \times 10^8 \text{ M}^{-1} \text{ s}^{-1}; k_{1b} = 1.4 \times 10^4 \text{ s}^{-1}).$$



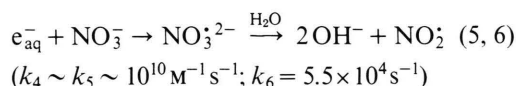
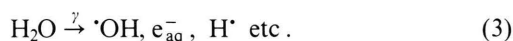
$$(k_2 = 1.0 \times 10^3 \text{ s}^{-1}).$$

In competition with these reactions NO<sub>2</sub> was found capable of oxidizing Fe(CN)<sub>6</sub><sup>4-</sup> at pH 7 [2] thus the oxidation-reduction potential of the NO<sub>2</sub>/NO<sub>2</sub><sup>-</sup> couple is probably well above 0.36 V, possibly in the order of 0.9 V [3]. The chemical behaviour of NO<sub>2</sub> in aqueous solutions containing organic compounds has however not been explored up to date, although NO<sub>2</sub> is supposed to be a highly deleterious agent in biological systems [4]. In the present study, using Gly-Tyr as model compound, it is shown that NO<sub>2</sub>-induced oxidation of tyrosine in dilute aqueous solution competes favourably with the natural decay of NO<sub>2</sub> by the reactions (1) and (2).

## Experimental

NO<sub>2</sub> was generated by  $\gamma$ -irradiation of deaerated aqueous solutions containing 10<sup>-2</sup> M NaNO<sub>2</sub> and

5 × 10<sup>-2</sup> M NH<sub>4</sub>NO<sub>3</sub> at pH > 8 (unbuffered). The system involves reactions (3) to (6), [1, 5–7]



and avoids reactions of e<sub>aq</sub><sup>-</sup> with O<sub>2</sub> and NO<sub>2</sub><sup>-</sup> [6]. The yield of NO<sub>2</sub><sup>•</sup> is Y(NO<sub>2</sub><sup>•</sup>) ~ 0.57 μM/Gy (G(NO<sub>2</sub><sup>•</sup>) = 5.5 per 100 eV)<sup>1</sup>; reactions of NO<sup>•</sup>, formed by interaction of H<sup>•</sup> with NO<sub>2</sub><sup>-</sup> [8], were not considered in view of the low yield (~ 0.06 μM/Gy).

Solutions were prepared shortly before each experiment with redistilled water, Gly-Tyr from Serva (Heidelberg) and A.R. grade inorganic chemicals. After deaeration, by 30 min bubbling with N<sub>2</sub> gas, the solutions were irradiated in the closed test tube at ambient temperature, using a 35 Gy/min <sup>60</sup>Co- $\gamma$ -source ("220 Gammacell", Atomic Energy of Canada Ltd.).

## Results and Discussion

$\gamma$ -Irradiation of the deaerated NO<sub>2</sub><sup>-</sup>/NO<sub>3</sub><sup>-</sup> system (see Experimental) in the absence of the model compound Gly-Tyr gave no detectable changes in the absorption spectra, indicating that NO<sub>2</sub> reverts quantitatively to the parent ions by the reactions (1) and (2). When Gly-Tyr was added to the NO<sub>2</sub><sup>-</sup>/NO<sub>3</sub><sup>-</sup> system, stable products were formed upon  $\gamma$ -irradiation with characteristic pH-dependent absorption

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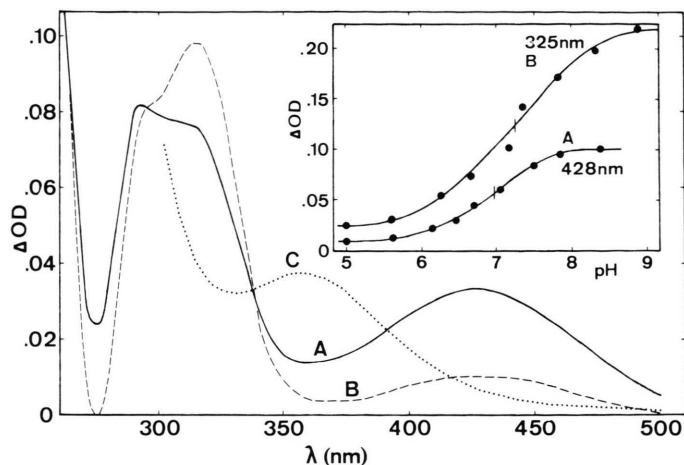


Fig. 1. Absorptions detected after  $\gamma$ -irradiation ( $D = 90$  Gy) of deaerated aqueous solutions containing 10 mM  $\text{NaNO}_2$  and 50 mM  $\text{NH}_4\text{NO}_3$  at pH 8.4 (unbuffered), (A) in the presence of 0.25 mM Gly-Tyr, (B) in the presence of 1 mM Gly-Tyr. All absorbances were measured against the corresponding unirradiated solution, using 1 or 5 cm cells; the optical densities ( $\Delta\text{OD}$ ) in the spectra A and B are normalized to 1 cm optical path. The pH dependences (insert), obtained by 1:1 dilution of the irradiated solutions A (428 nm) and B (325 nm) with buffers, refer to 5 cm optical path; the spectrum C is that of the diluted (1:1) solution A at pH 5.7.

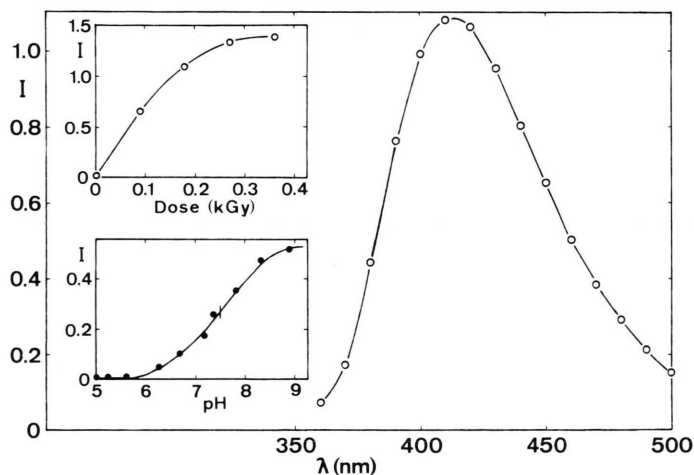


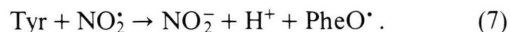
Fig. 2. Fluorescence of products detected after  $\gamma$ -irradiation of a deaerated solution containing 10 mM  $\text{NaNO}_2$ , 50 mM  $\text{NH}_4\text{NO}_3$  and 1 mM Gly-Tyr at pH 8.4. The fluorescence was excited at 325 nm and the intensity ( $I$ ) was measured relative to a 0.5 mM Na-salicylate reference solution. The spectrum refers to  $D = 180$  Gy, and the pH dependence refers to  $D = 90$  Gy and subsequent dilution (1:1) with buffers (cf. Fig. 1B). 410 nm fluorescence intensities are given in the inserts.

and fluorescence spectra, as shown in Figs. 1 and 2. The Gly-Tyr concentration in these experiments was sufficiently low to exclude reactions of  $\cdot\text{OH}$  and  $\text{e}_{\text{aq}}^-$  with the peptide. It is evident thus that  $\text{NO}_2^-$ , generated by the reactions (4) to (6), is capable of reacting with Gly-Tyr in competition with the natural  $\text{NO}_2^-$  decay by reactions (1) and (2), even at low peptide concentrations (0.25 mM).

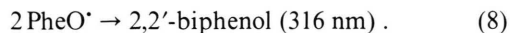
In an attempt at product identification we recall that phenoxyl radicals formed by 1-electron oxidation of Gly-Tyr (e.g. by  $\text{N}_3$ ) efficiently dimerize to form the Gly-Tyr 2,2'-biphenol [9], which deprotonates at pH  $\sim 7.4$ ; the alkaline form of 2,2'-biphenol (not the acid form) absorbs with a maximum at 316 nm ( $\epsilon_{316} = 5790 \text{ M}^{-1} \text{ cm}^{-1}$ ) [10] and exhibits a strong fluorescence with a peak at 410 nm. The characteristic 316 nm absorption and

the 410 nm fluorescence (excited at 325 nm) is clearly seen at pH 8.4 in Fig. 1B and Fig. 2, respectively, and it was also confirmed that this species disappears at lower pH with  $\text{pK} \sim 7.4$  (see OD (325 nm) and  $I$  (410 nm) titration curves in the Figures). The 2,2'-biphenol does, however, not exhibit absorption peaks at 290 and 428 nm, as indicated particularly in the spectrum at low peptide concentration (Fig. 1A). The 290/428 nm absorption is characteristic for nitro-tyrosine, and an extinction coefficient of  $\epsilon_{428}(\text{Nitro-Tyr}) = 4100 \text{ M}^{-1} \text{ cm}^{-1}$  has been reported [11]. Nitro-tyrosine deprotonates at  $\text{pK} \sim 7$ , with an accompanying shift in the absorption maximum to 360 nm [11]. This behaviour is clearly demonstrated (Fig. 1) by the 428 nm titration curve and the spectrum C at pH 5.7.

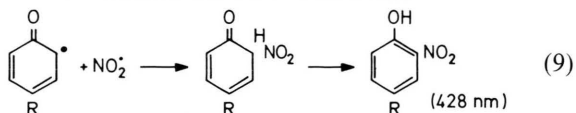
The above observations suggest that NO<sub>2</sub><sup>•</sup>, like N<sub>3</sub><sup>•</sup> [9], is capable of electron (or H-atom) abstraction from tyrosine to form phenoxyl radicals (PheO<sup>•</sup>),



About 62% of the phenoxyls dimerize in the absence of other reagents to form 2,2'-biphenols [9],



Formation of nitro-tyrosine can conceivably arise from interactions of PheO<sup>•</sup> with NO<sub>2</sub>,



Reaction (8), *i.e.* the 316 nm species, predominates at high tyrosyl concentration (Fig. 1 B) and reaction (9), *i.e.* the 428 nm species, contributed particularly at low tyrosyl concentration (Fig. 1 A), where reaction (7) is slower.

From Fig. 1 the product yields,  $Y = \text{OD}/(\epsilon \cdot D)$ , can be roughly estimated. By assuming that OD(428) = 0.036 (Fig. 1 A) is due to nitro-Tyr only, and OD(316) = 0.099 (Fig. 1 B) is pertinent to 2,2'-biphenol in 85% (since nitro-Tyr contributes), we find  $Y(\text{nitro-Tyr}) \sim 0.1 \mu\text{M/Gy}$  at 0.25 mM Gly-Tyr, and  $Y(2,2'\text{-biphenol}) \sim 0.16 \mu\text{M/Gy}$  at 1 mM Gly-Tyr.

Since two NO<sub>2</sub><sup>•</sup> radicals ( $Y(\text{NO}_2^\bullet) \sim 0.57 \mu\text{M/Gy}$ ) are required for each tyrosine dimer, this means that 56% of the NO<sub>2</sub><sup>•</sup> end up in 2,2'-biphenol at 1 mM Gly-Tyr. Comparing this value with the phenoxyl  $\rightarrow$  2,2'-biphenol yield (62%) [9], it can be concluded that NO<sub>2</sub><sup>•</sup> almost quantitatively oxidizes tyrosine under these conditions.

## Conclusions

The results presented reveal that NO<sub>2</sub><sup>•</sup> can act as a strong one-electron oxidant in aqueous environment; oxidation of the tyrosyl model compound competes favourably, even at low concentrations, with the disproportionation of NO<sub>2</sub><sup>•</sup> in water. It can be anticipated therefore that NO<sub>2</sub><sup>•</sup> is capable also of inactivating proteins and other constituents of living cells such as thiols. Such direct action of NO<sub>2</sub><sup>•</sup> may be generally involved in the deleterious effects initiated by NO<sub>2</sub><sup>•</sup> in biological systems. It should be noted also that oxidation (reaction (7)), as compared to disproportionation (reaction (2)), generates up to twice the yield of harmful nitrite.

## Acknowledgements

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- [1] M. Grätzel, A. Henglein, J. Lilie, and G. Beck, *Ber. Bunsenges. phys. Chem.* **73**, 646 (1969). Since an incorrect relation ( $k_{1b} = K \cdot k_{1f}$ ) was used by these authors, we have quoted  $k_{1b} = K \cdot 2 k_{1f}$ .
- [2] M. Ottolenghi and J. Rabani, *J. Phys. Chem.* **72**, 593 (1968).
- [3] H. Pick, *Z. Elektrochem.* **26**, 182 (1920).
- [4] R. L. Heath, *Ann. Rev. Plant Physiol.* **31**, 395 (1980).
- [5] Farhataziz and A. B. Ross, *NSRDS-NBS 59* (U.S. Government Printing Office, Washington 1977).
- [6] M. Anbar, M. Bambenek, and A. B. Ross, *NSRDS-NBS 43* (U.S. Government Printing Office, Washington, 1973).
- [7] M. Grätzel, A. Henglein, and S. Taniguchi, *Ber. Bunsenges. phys. Chem.* **74**, 292 (1970).
- [8] M. Anbar, Farhataziz, and A. B. Ross, *NSRDS-NBS 51* (U.S. Government Printing Office, Washington 1975).
- [9] W. A. Prütz, J. Butler, and E. J. Land, *Int. J. Radiat. Biol.* **44**, 183 (1983).
- [10] A. J. Gross and I. W. Sizer, *J. Biol. Chem.* **234**, 1611 (1959).
- [11] M. Sokolovsky, J. F. Riordan, and B. L. Vallee, *Biochemistry* **5**, 3582 (1966).